

0960-894X(93)E0098-L

SYNTHESIS AND BIOLOGICAL ACTIVITIES OF FLUORINE-SUBSTITUTED (-)-INDOLACTAM-V, THE CORE STRUCTURE OF TUMOR PROMOTER TELEOCIDINS

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Abstract: To investigate the conformation-activity relationship, new indolactam derivatives with a fluorine atom at position 5 or 6 were synthesized. (-)-5-Fluoroindolactam-V (2) existing mainly as the sofa conformer in solution was far less biologically active than (-)-6-fluoroindolactam-V (3) existing mainly as the twist conformer. This suggests that the sofa conformation of (-)-indolactam-V (1) is not a biologically active form.

Teleocidins (for example, teleocidin A-1 and teleocidin B-4)¹⁻³⁾ are potent skin tumor promoters⁴⁾ produced by actinomycetes. Teleocidins and their core structure (-)-indolactam-V (1)^{5,6)} exist as two stable conformers, a twist and a sofa form, in solution at room temperature.⁷⁾ Although recent structure-activity studies using a number of indolactam derivatives⁸⁻¹³⁾ have revealed the essential structural factors for tumor-promoting activity, the active conformation of teleocidins remains unclear. To solve this problem, conformationally restricted analogues of teleocidins are indispensable. We have previously reported that 5-substituted indolactam derivatives such as (-)-5-acetyl- and (-)-5-chloroindolactam-V exist exclusively as the sofa conformer.¹⁴⁾ However, the biological activities of these derivatives are expected to reflect both the conformational change and the steric effect at position 5 of the indole ring. To estimate more exactly the activity of the sofa conformer, it is necessary to minimize the steric, electronic, and hydrophobic effects of the substituent at position 5. The most promising candidate is (-)-5-fluoroindolactam-V (2). This communication describes the synthesis and biological activities of (-)-5-fluoroindolactam-V (2) along with those of (-)-6-fluoroindolactam-V (3) as a positive control.

Since most fluorinating agents are explosive, toxic, or unstable materials which require special equipment and techniques, and since the locus of electrophilic substitution reactions on (-)-indolactam-V (1) is position 7,¹⁴) we abandoned the route of the direct fluorination of 1. We have recently shown that the microbial conversion of seco-compounds (N-methyl-L-amino acidyl-L-tryptophanol) of indolactams is a simple and convenient method to prepare various indolactam analogues.¹⁵ Therefore, we tried to expand this microbial conversion to substituted N-methyl-L-tryptophanols; 2 and 3 were synthesized from the corresponding fluorinated DL-tryptophans using the microbial conversion as a key step.

Figure 1 Naturally-occurring teleocidin-related compounds.

$$R_1$$
 OH R_2 N R_2 N R_3 R_4 R_4 R_5 R_1 R_2 R_4 R_5 R_7 R_8 R_8 R_8 R_8 R_8 R_8 R_8 R_9 R_9

a) SOCl₂, McOH, reflux b) LiAlH₄, THF, reflux c) Boc-N-methyl-L-valine, DCC, HONSu, THF d) i) IM HCl/AcOH ii) NaOH, H₂O, McOH e) S. blastmyceticum NA34-17, 36hr

Figure 2 Synthesis of fluorine-substituted indolactams (2 and 3).

Table 1. Several characteristic signals of the ¹H NMR spectra of (-)-indolactam-V and its fluorine-substituted analogues in CDCl₃ (400MHz)

Posi	tion	δ (multiplicity, J in Hz)							
	(-)-Indolactam	a-V (1) (-)	-5-Fluoroindolactam-V (2) (-)	(-)-6-Fluoroindolactam-V (3)					
	twist conformera	sofa conformer ^a	sofa conformer ^b	twist conformer ^c					
1	8.03 (br.s)	8.35 (br.s)	8.27 (br.s)	7.98 (br.s)					
10	7.36 (br.s)	4.74 (d, <i>J</i> =11.6)	4.72 (d, <i>J</i> =11.0)	7.18 (br.s)					
12	4.40 (d, <i>J</i> =10.1)	2.99 (d, <i>J</i> =11.0)	2.90 (d, <i>J</i> =11.0)	4.40 (d, J=10.4)					
14	3.57 (dd, J=11.3, 8.5)	3.41 (dd, <i>J</i> =11.0, 7.0)	3.43 (m)	3.57 (dd, <i>J</i> =11.3, 8.5)					
	3.74 (dd, <i>J</i> =11.3, 3.7)			3.75 (dd, <i>J</i> =11.3, 3.7)					
15	2.60 (m)	2.40 (m)	2.46 (m)	2.59 (m)					
16	0.63# (d, <i>J</i> =6.7)	0.94+ (d, <i>J</i> =6.4)	0.94§ (d, <i>J</i> =6.7)	0.64* (d, <i>J</i> =6.7)					
17	0.93# (d, <i>J</i> =6.4)	1.25+ (d, <i>J</i> =6.4)	1.26§ (dd, $J=6.7, 3.4$)	0.93* (d, <i>J</i> =6.4)					
18	2.92 (s)	2.75 (s)	2.76 (s)	2.88 (s)					

asofa:twist = 1:4.5 (0.033M, 27°C). bsofa > 95% (0.013M, 27°C). csofa:twist = 1:15 (0.015M, 27°C).

The seco-compounds, 5-fluoro- or 6-fluoro-N-methyl-L-valyl-L-tryptophanol (4, 5), ¹⁶⁾ were synthesized from Boc-N-methyl-L-valine and DL-5- or DL-6-fluorotryptophan by the method reported previously. ¹⁵⁾ Compounds 4 and 5 were then converted to 2 and 3¹⁷⁾ at 6.7% and 8.1% yield, respectively, by Streptoverticillium blastmyceticum NA34-17 (Fig. 2). ^{6,18)} This is the first report of synthesis of 6-substituted indolactams. Since the orientation of the electrophilic substitution reaction on 1 is position 7, ¹⁴⁾ this microbial conversion would be a simple and convenient method to prepare a series of 6-substituted indolactams; in fact, we have also succeeded to synthesize (-)-6-methylindolactam-V by a similar method (data not shown). The ¹H NMR spectra indicated that 2 and 3 existed mainly as the sofa and the twist conformer, respectively (Table 1). This was determined by several characteristic signals of the two conformers. ⁷⁾ The sign of $[\alpha]_D$ of 2 was opposite to that of 3, ¹⁷⁾ indicating the existence of a large difference in the tertiary structure between them.

^dAssignments bearing the same symbol may be reversed.

Table 2. B	iological activities of	(-)-indolactam-V	and fluorine-substituted analogues
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Compound	Inhibition of specific [3H]TPA binding ^a	Incorporation of ³² P _i into phospholipids of HeLa cells ^b					
•		relative cpm/mg of protein					
	ID ₅₀ (log 1/M)	10 ⁻⁸ M	10 ⁻⁷ M	10-6M	10 ⁻⁵ M	10 ^{-4 5} M	
DMSO (control)°		1.00 (0.06) ^d					
(-)-Indolactam-V (1)	5.97 (0.10)	1.04 (0.05)	1.36 (0.22)	4.69 (0.49)	7.82 (0.37)	NTe	
(-)-5-Fluoro- indolactam-V (2)	4.41 (0.14)	NT	1.29 (0.07)	1.28 (0.06)	3.57 (0.33)	5.32 (0.38)	
(-)-6-Fluoro- indolactam-V (3)	5.85 (0.01)	1.18 (0.13)	1.78 (0.01)	7.43 (0.20)	6.87 (0.00)	NT	

^aThis inhibition assay was carried out by the cold acetone filter method. ¹⁹⁾ ^bThis assay was done by the method reported previously²⁰⁾ with slight modifications. ¹³⁾ ^{c0.2%} DMSO. ^dStandard deviation. ^eNot tested.

The tumor-promoting activity of these compounds was examined by two *in vitro* bioassays: binding affinity to the 12-O-tetradecanoylphorbol-13-acetate (TPA) receptor in the mouse epidermal particulate fraction and stimulation of the incorporation of radioactive inorganic phosphate ($^{32}P_i$) into phospholipids of HeLa cells. Table 2 summarizes the results of these assays. The binding affinity of (-)-5-fluoroindolactam-V was about 1/40 of that of (-)-indolactam-V and about 1/30 of that of (-)-6-fluoroindolactam-V. (-)-Indolactam-V had a maximum increase of $^{32}P_i$ incorporation at ^{10-5}M and (-)-6-fluoroindolactam-V at ^{10-6}M . By contrast, (-)-5-fluoroindolactam-V did not maximally increase the $^{32}P_i$ incorporation even at $^{10-4.5}M$. These two biological activities correlated very well for each derivative.

The sofa form has been hypothesized to be very close to the active conformation of teleocidins from the results of the superposition of teleocidins and phorbol esters²¹⁾ and molecular dynamics calculations.²²⁾ However, our present findings suggest that the sofa conformation of (-)-indolactam-V is far from the biologically active form because (-)-5-fluoroindolactam-V existing mainly as the sofa form was biologically nearly inactive. A similar conclusion has been recently reported by Kozikowski et al.²³⁾ and Ohno et al.²⁴⁾ using newly designed benzolactams, indolactam analogues without pyrrole moiety. A high level of the activities of (-)-6-fluoroindolactam-V existing mainly as the twist form seems to indicate that the active conformation is the twist form. However, according to Kawai et al.²²⁾ at least ten conformations of indolactams should be considered as candidates for the active conformation because of the low energy barrier among them. In conclusion, the present study excluded the possibility of the sofa conformer as the biologically active form. Further synthetic studies of other conformationally restricted analogues are necessary to determine the active conformation of teleocidins.

Acknowledgement

We thank Drs H. Hayashi and M. Arai of the Department of Agricultural Chemistry at the University of Osaka Prefecture for cultivating the micro-organisms; Mr. R. Imamura of the Faculty of Science at Kyoto University for the ¹H NMR measurements. This work was supported in part by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science and Culture of Japan (K.I.).

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- 5-Fluoro-N-methyl-L-valyl-L-tryptophanol (4): [α]_D -17.0° (c=1.06, MeOH, 23°C). UV λ_{max} (MeOH) nm (ε): 287(4,200), 222.5 (16,100). ¹H NMR δ (CD₃OD, 0.10M) ppm: 0.85 (3H, d, J=7.0Hz), 0.87 valyl-L-tryptophanol (5): $[\alpha]_D$ -35.6° (c=0.96, MeOH, 21°C). UV λ_{max} (MeOH) nm (ϵ): 283 (3,900), 220 (22,400). ¹H NMR & (CD₃OD, 0.06M) ppm: 0.85 (3H, d, J=6.4Hz), 0.87 (3H, d, J=6.4Hz), 1.78 (1H, m), 2.04 (3H, s), 2.66 (1H, d, J=6.4Hz), 2.87 (1H, dd, J=14.7, 8.5Hz), 3.05 (1H, dd, J=14.7, 6.1Hz), 3.57 (1H, dd, J=11.0, 5.5Hz), 3.61 (1H, dd, J=11.0, 5.5Hz), 4.29 (1H, m), 6.78 (1H, ddd, J=11.0, 5.5Hz), 3.61 (1H, dd, J=11.0, 5.5Hz), 4.29 (1H, m), 6.78 (1H, ddd, J=11.0, 5.5Hz), 3.61 (1H, dd, J=11.0, 5.5Hz), 4.29 (1H, m), 6.78 (1H, ddd, J=11.0, 5.5Hz), 4.29 (1H, m), 6.78 (1H, ddd, J=11.0, 5.5Hz), 4.29 (1H, dd, J=9.8, 8.5, 2.4Hz), 6.99 (1H, dd, J=9.8, 2.4Hz), 7.08 (1H, s), 7.58 (1H, dd, J=8.5, 5.2Hz). HR-
- EIMS m/z: 321.1823 (M+, calcd. for $C_{17}H_{24}N_3O_2F$, 321.1852). 17. (-)-5-Fluoroindolactam-V (2): $[\alpha]_D$ +71.7° (c=0.10, MeOH, 26°C). UV λ_{max} (MeOH) nm (ϵ): 299 (7,600), 289 (8,100), 223.5 (25,800). ¹H NMR δ (CDCl₃, 0.013M, sofa > 95%) ppm: sofa conformer; 0.94 (3H, d, J=6.7Hz), 1.26 (3H, dd, J=6.7, 3.4Hz), 2.46 (1H, m), 2.76 (3H, s), 2.82 (1H, dd, J=14.7, 1.5Hz), 2.90 (1H, d, J=11.0Hz), 3.10 (1H, dd, J=14.3, 4.3Hz), 3.43 (2H, m), 4.47 (1H, m), 4.72 (1H, d, J=11.0Hz), 7.01 (1H, dd, J=10.1, 8.6 Hz), 7.07 (1H, d, J=2.4Hz), 7.19 (1H, dd, J=8.6, 4.0Hz), 8.27 (1H, br.s). HR-EIMS m/z: 319.1693 (M*, calcd. for $C_{17}H_{22}N_3O_2F$, 319.1696). (-)-6-Fluoroindolactam-V (3): $[\alpha]_D$ -386.0° (c=0.11, MeOH, 18°C). UV λ_{max} (MeOH) nm (ϵ): 292 (11,000), 227 (28,000). 1H NMR δ (CDCl₃, 0.015M, sofa:twist = 1:15) ppm: twist conformer; 0.64 (3H, d, 1.67 (Hz), 0.03 J=6.7Hz), 0.93 (3H, d, J=6.4Hz), 2.59 (1H, m), 2.88 (3H, s), 3.01 (1H, dd, J=17.4, 3.7Hz), 3.15 (1H, br.d, J=17.4Hz), 3.57 (1H, dd, J=11.3, 8.5Hz), 3.75 (1H, dd, J=11.3, 3.7Hz), 4.22 (1H, br.s), 4.40 (1H, d, J=10.4Hz), 6.28 (1H, dd, J=12.2, 2.1Hz), 6.57 (1H, dd, J=8.5, 2.1Hz), 6.86 (1H, s), 7.18 (1H, br. s), 7.98 (1H, br. s). HR-EIMS m/z; 319.1683 (M+, calcd. for C₁₇H₂₂N₃O₂F, 319.1696). 18. Irie, K.; Hagiwara, N.; Koshimizu, K.; Hayashi, H.; Murao, S.; Ito, Y. Agric. Biol. Chem. 1985, 49,

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